

## ERASMUS WORK PLACEMENT

| EMPLOYER INFORMATION                              |  |
|---|--|
| Organization/ Institution/ Enterprise/ University | Dpt. Of Chemistry, Faculty of Experimental Sciences.<br>University of Huelva |
| Website   | <a href="http://www.uhu.es/algatech/">http://www.uhu.es/algatech/</a>        |

| CONTACT DETAILS                   |  |
|-----------------------------------|--|
| Contact people for this placement | Inés Garbayo Nores                                 |
| Department                        | Chemistry  |
| Direct telephone number           | +34 959 21 99 53                                   |
| E-mail addresses                  | <a href="mailto:garbayo@uhu.es">garbayo@uhu.es</a> |

| PLACEMENT INFORMATION      |  |
|----------------------------|--|
| Function of the Department | Laboratory of Biotechnology of Algae   |
| Description of activities  | <p>Our lab goal is to provide students a variety of techniques in the field of bioproduction of antioxidants from microalgae. The aim of our research is to define conditions which enhance biosynthesis, promote accumulation and address full biocompatible extraction of carotenoids antioxidants from different microalgae (Chlamydomonas acidophila, Nannochloropsis gaditana, Dunaliella salina among others).</p> <p>We always encourage students to get involved in our projects under supervision and then focus in on ones that are the most interest to them.</p> <p>Laboratory equipment and techniques used :</p> <p>Cell microalgae cultures: Growth chamber for photosynthetic microorganisms with controlled temperature and different type and quality of light. Optic microscopy to observe cellular growth. Hoods with laminar flux. Autoclave for sterilization processes.</p> <p>Protein purification: Refrigerated chamber (4°C). Purification systems (chromatographic columns, peristaltic pumps, collector of fractions). Chromatographic system for protein purification (FPLC).</p> <p>Separation and quantification of target compounds: Chromatography techniques (HPLC with fluorescence and UV-vis detector). UV-visible spectroscopy</p> |

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|-------------------------|--|
|                         | Continuous long-term cultures: 5-liters bioreactor with pH, T <sup>a</sup> and oxygen probe. Software for an automatic continuous control of the reactor.<br>Determination of cellular viability: Oxygen Clark |
| Location                | Huelva, Spain  |
| Duration                | 3-12 months  |
| Working hours per week: | Depends  |
| Start Date:             | Rolling  |
| Accommodation:          | Not specified  |
| Application Procedure:  | Send CV and cover letter to Inés Garbayo   |

### COMPETENCES, SKILLS and EXPERIENCE REQUIREMENTS

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| Languages and level of competence required   | Spanish, English, or French (Spanish not required) |
| Computer skills and level of skills required | N/A  |

Detailed programme of the training at the **Laboratory of Biotechnology of Algae**.  
Dpt. of Chemistry. Faculty of Experimental Sciences. University of Huelva.

#### Training of 3 months

**Week 1** Bibliographical review

**Week 2** Optimization of culture medium and growth of *Chlamydomonas acidophila*, *Nannochloropsis gaditana*, *Dunaliella salina* and determination of cellular viability with Oxygen Clark electrode and PAM system for determination and quantification of chlorophyll.

**Week 3** HPLC techniques for lutein and  $\beta$ -carotene separation. UV-visible spectroscopy

**Week 4-7** Culture and accumulation of lutein in *C. acidophila*/ *N. gaditana*/ *D. salina* under different nutritional stress conditions (nitrogen, sulfur, UV radiation, pH, heavy metals). Proposal of best conditions to produce highest amounts of lutein.

**Week 8-10** Proposal of long-term processes with lab scale reactors for continuous production of lutein in bioreactors. 5-liters bioreactor with pH, T<sup>a</sup> and oxygen probe. Software for an automatic continuous control of the reactor.

**Week 11-12** Final analysis and discussion of results. Proposal and final report.

## Training of 6 months

**Week 1** Bibliographical review

**Week 2-3** Optimization of culture medium and growth of *Chlamydomonas acidophila*, *Nannochloropsis gaditana*, *Dunaliella salina* and determination of cellular viability with Oxygen Clark electrode and PAM system for determination and quantification of chlorophyll.

**Week 4-5** HPLC techniques for lutein and  $\beta$ -carotene separation. UV-visible spectroscopy

**Week 6-9** Culture and accumulation of lutein in *C. acidophila*/ *N. gaditana*/ *D. salina* under different nutritional stress conditions (nitrogen, sulfur, UV radiation, pH, heavy metals). Proposal of best conditions to produce highest amounts of lutein.

**Week 10-13** Proposal of long-term processes with lab scale reactors for continuous production of lutein in bioreactors. 5-liters bioreactor with pH, T<sup>a</sup> and oxygen probe. Software for an automatic continuous control of the reactor.

**Week 14-16** Electronic Microscopic studies of ultrastructural changes induces by oxidative stress in microalgae cells of *Chlamydomonas*, *Dunaliella* and *Nannochloropsis*.

**Week 17-21** Microalgae cultures in *outdoor* reactors using best conditions obtained at lab scale reactors. Production of lutein/ $\beta$ -carotene using optimal conditions obtained in weeks 6-9 and 10-13.

**Week 22-24** Final analysis and discussion of results. Proposal and final report.